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### Easy access to optically active Hagemann's esters

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**Abstract**—The synthesis of optically active Hagemann's esters was investigated. The starting materials in this approach were enamino esters (R,Z)-8, prepared through the condensation of keto ester 6 with (R)-1-phenylethylamine 7. Michael addition reaction of the enamino esters (R,Z)-8 with methyl vinyl ketone gave the expected adducts 10 with good e.e.s of 93–96%. Subsequent annulation of the adducts furnished optically active Hagemann's esters. © 2001 Elsevier Science Ltd. All rights reserved.

#### 1. Introduction

Commercially available Hagemann's ester 1 and its analogues have been extensively used in the synthesis of a variety of complex molecules, including terpenes, alkaloids, taxane core, steroids and flavones. As a part of our program directed toward synthesising new chiral building blocks, the enantiopure four-substituted Hagemann's esters 2 have recently attracted our attention (Scheme 1).

Although several racemic Hagemann's esters **2** have been previously prepared,<sup>2</sup> to our knowledge only one stereoselective approach to such a compound, (S)-**2** ( $R = CH_3$ ;  $R' = C_2H_5$ ), based on the microbial reduction of a  $\beta$ -keto ester,<sup>3</sup> has been disclosed so far.

We planned to elaborate such molecules through an efficient enantioselective methodology we have developed based on the asymmetric Michael addition reaction using chiral  $\beta$ -enamino esters. Thus, condensation of chiral enamino-esters 3 (derived from cyclic or acyclic  $\beta$ -keto esters and enantiopure 1-phenylethylamine) to electron-deficient alkenes 4 under neutral conditions furnished, after hydrolytic work-up,  $\beta$ -keto esters 5, in moderate yields and excellent enantiomeric excesses (e.e.s) (Scheme 2).

Herein, we report a short, efficient enantioselective approach to various Hagemann's esters of type 2 by

applying this methodology. These compounds were obtained in good yields, and with excellent e.e.s of 93–96%.

#### 2. Results and discussion

The enamino esters **8** were first prepared from reaction between the 2-substituted acetoacetate **6** and enantiomerically pure (R)-1-phenylethylamine **7** over 12 hours in toluene at reflux in the presence of catalytic p-TsOH. The (R)-esters were isolated in 72–86% purified yield. The (Z)-geometry in these compounds is secured by intramolecular hydrogen bonding.

Scheme 1.

Ph. Me 
$$i:$$
 EWG  $\frac{1}{4}$  EWG  $\frac{1}{4}$   $\frac{1}$ 

Scheme 2.

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a: R = Me, R' = Et b: R = Et, R' = Et c: R = Allyl, R' = Me d: R = Bn, R' = Me

#### Scheme 3.

Addition of enamino ester (R)-8 to methyl vinyl ketone 9 required the presence of 1 equiv. of zinc chloride. This condensation, which was carried out in THF for 1 hour at 0°C and quenched with 20% aqueous AcOH, furnished  $\alpha$ , $\alpha$ -disubstituted  $\beta$ -keto esters 10 in 67–75% yield (Scheme 3).

The e.e.s of **10** (93–96%) were determined by <sup>1</sup>H NMR spectroscopy in the presence of the chiral shift reagent Eu(hfc)<sub>3</sub>. The sense of induction of this Michael addition was deduced from mechanistic considerations (vide infra) and was unequivocally determined through the correlation of Hagemann's ester **2a** to the known derivative (S)-**2a**.<sup>3</sup>

As previously reported,<sup>2</sup> catalytic piperidinium acetate effects cyclisation of **10** furnishing mainly Hagemann's esters **2** in 65–79% yields (Scheme 4).

It is of interest to note that the stereochemical outcome observed in the previous Michael reactions using acyclic enamino esters 8 follows the general mechanism established in this series.<sup>5</sup> In accordance with this mechanism, the reaction proceeds through the 'aza-ene'-like transition state 11, in which the N-H proton of the enamino ester is transferred to the α-carbon atom of the electrophilic alkene concertedly with the creation of the C-C bond. This requires a synclinal arrangement of the two reactants, as shown in the corresponding compact approach 12. According to such a model, alkylation takes place predominantly anti to the bulky phenyl ring of the chiral amine moiety portrayed in its energetically preferred conformation, minimising A<sup>1,3</sup>-type strain, with the C-H bond more or less eclipsing the enamine ring. This accounts for the absolute configuration in adducts 10 (Scheme 5).

Studies directed at the utilisation of Hagemann's esters **2c** and **2d** as chiral building blocks are currently under investigation in our laboratory.

### 3. Experimental

### 3.1. General

Melting points were recorded on a Kofler bench. Infrared (IR) spectra were obtained on a Perkin-Elmer

881 as neat films between NaCl plates or KBr pellets. The <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Bruker AC 200 P (200 and 50 MHz, for <sup>1</sup>H and <sup>13</sup>C, respectively) in CDCl<sub>3</sub>. Recognition of methyl, methylene, methine and quaternary carbon nuclei in <sup>13</sup>C NMR spectra rests on the J-modulated spin echo sequence. Optical rotations were measured at 20°C on a Polax L polarimeter in a 1 dm cell at 589 nm. Analytical thinlayer chromatography was performed on Merck silica gel 60F<sub>254</sub> pre-coated plates. All solvents were purified before use. Tetrahydrofuran (THF) was distilled from sodium-benzophenone ketyl. Methanol was dried over magnesium and distilled. Toluene was distilled from calcium hydride. All reactions involving air- or watersensitive compounds were routinely conducted in flamedried glassware under positive pressure of nitrogen. Organic extracts were dried over anhydrous MgSO<sub>4</sub>. Compounds obtained from commercial suppliers were used without further purification. All elemental analyses were performed by the Service de Microanalyse, Centre d'Etudes Pharmaceutiques, Châtenay-Malabry, France, with a Perkin-Elmer 2400 analyser.

## 3.2. General procedure for the preparation of enamino esters

To a solution of keto ester 6 (0.05 mol) and p-toluene-sulfonic acid (catalytic) in anhydrous toluene (100 mL) was added R-(+)-1-phenylethylamine (0.0055 mol). The mixture was stirred under reflux for 14 h with azeotropic removal of water using a Dean–Stark trap. The solution was cooled to  $20^{\circ}$ C, concentrated in vacuo and the residue purified by distillation.

a: R = Me, R' = Et b: R = Et, R' = Et c: R = Allyl, R' = Me d: R = Bn, R' = Me

Scheme 4.

Scheme 5.

**3.2.1.** (*R*)-2-Methyl-3-(1-phenylethylamino)-but-2-enoic acid ethyl ester 8a. Oil; yield 86%; bp (0.1 mmHg):  $135^{\circ}$ C;  $[\alpha]_{D} = -298$  (c = 5.5, CHCl<sub>3</sub>); IR (neat, cm<sup>-1</sup>): 3244, 1646, 1603, 1592, 1450; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  1.3 (t, J = 7.0 Hz, 3H), 1.5 (d, J = 7.0 Hz, 3H), 1.75 (s, 3H), 4.19 (q, J = 7.0 Hz, 2H), 4.6 (quint, J = 7.0 Hz, 1H), 7.2 - 7.4 (m, 5H), 9.6 (d, J = 7.0 Hz, 1H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  172.0 (C), 159.0 (C), 142.5 (C), 128.9 (2 CH), 127.2 (CH), 125.5 (2 CH), 87.5 (C), 58.6 (CH<sub>2</sub>), 53.0 (CH), 25.1 (CH<sub>3</sub>), 15.6 (CH<sub>3</sub>), 14.5 (CH<sub>3</sub>), 12.4 (CH<sub>3</sub>).

**3.2.2.** (*R*)-2-Ethyl-3-(1-phenylethylamino)-but-2-enoic acid ethyl ester 8b. Syrup; yield 72%; bp (0.1 mmHg):  $140^{\circ}$ C;  $[\alpha]_{D} = -315.2$  (c = 2.85, CHCl<sub>3</sub>); IR (neat, cm<sup>-1</sup>): 3250, 2980, 1650, 1600; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  0.9 (t, J = 6.0 Hz, 3H), 1.38 (t, J = 7.0 Hz, 3H), 1.40 (d, J = 7 Hz, 3H), 1.78 (s, 3H), 2.20 (q, J = 6 Hz, 2H), 4.10 (q, J = 7 Hz, 2H), 4.60 (quint, J = 7.0 Hz, 1H), 7.10 - 7.30 (m, 5H), 9.65 (d, J = 7 Hz, 1H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  176.0 (C), 159.5 (C), 142.5 (C), 128.2 (2 CH), 126.8 (CH), 124.9 (2 CH), 95.0 (C), 59.4 (CH<sub>2</sub>), 52.4 (CH), 24.8 (CH<sub>3</sub>), 22.5 (CH<sub>2</sub>), 17.5 (CH<sub>3</sub>), 16.8 (CH<sub>3</sub>), 16.6 (CH<sub>3</sub>).

3.2.3. (*R*)-2-Allyl-3-(1-phenylethylamino)-but-2-enoic acid methyl ester 8c. Syrup; yield 75%; bp (0.1 mmHg):  $140^{\circ}$ C;  $[\alpha]_{D} = -387$  (c = 12.3, CHCl<sub>3</sub>); IR (neat, cm<sup>-1</sup>): 3230, 2900, 1648, 1600; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  1.25 (d, J = 6.0 Hz, 3H), 1.78 (s, 3H), 2.85–3.0 (m, 2H), 3.69 (s, 3H), 4.60 (quint, J = 6.0 Hz, 1H), 4.80–4.90 (m, 2H), 5.60–5.90 (m, 1H), 7.10–7.29 (m, 5H), 9.80 (d, J = 6.0 Hz, 1H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  176.0 (C), 160.6 (C), 145.5 (C), 138.3 (CH), 128.8 (2 CH), 127.0 (CH), 125.5 (2 CH), 113.0 (CH<sub>2</sub>), 90.1 (C), 53.2 (CH<sub>3</sub>), 50.5 (CH), 31.1 (CH<sub>2</sub>), 25.2 (CH<sub>3</sub>), 15.3 (CH<sub>3</sub>).

**3.2.4.** (*R*)-2-Benzyl-3-(1-phenylethylamino)-but-2-enoic acid methyl ester 8d. Syrup; yield 84%; bp (0.1 mmHg):  $150^{\circ}$ C;  $[\alpha]_{D} = -310$  (c = 1.4, CHCl<sub>3</sub>); IR (neat, cm<sup>-1</sup>): 3190, 1717, 1648; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  1.25 (d, 3H, J = 7.0 Hz), 1.70 (s, 3H), 3.48 (d, J = 14 Hz, 1H), 3.70 (d, J = 14 Hz, 1H), 3.80 (s, 3H), 4.60 (quint,

J=7 Hz, 1H), 7.00–7.40 (m, 5H), 9.90 (d, J=7.0 Hz 1H);  $^{13}$ C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  171.13 (C), 158.8 (C), 142.2 (C), 137.5 (C), 90.1 (C), 129.2 (2 CH), 128.4 (2 CH), 128.1 (2 CH), 127.3 (2 CH), 126.7 (CH), 125.5 (CH), 52.7 (CH), 50.8 (CH<sub>3</sub>), 30.7 (CH<sub>2</sub>), 24.9 (CH<sub>3</sub>), 18.6 (CH<sub>3</sub>).

# 3.3. General procedure for the addition of methyl vinyl ketone

Methyl vinyl ketone (0.055 mol) was added to a solution of ZnCl<sub>2</sub> (catalytic) in anhydrous toluene (40 mL). The mixture was stirred for 1 h at 0°C. A solution of enamino ester 8 (0.05 mol) in anhydrous toluene (5 mL) was added and the mixture was stirred at 0°C for 2 h. A 10% aqueous acetic acid solution was then added (30 mL) and the resulting mixture was stirred for 2 h. The aqueous phase was saturated with NaCl and extracted with ethyl acetate (3×50 mL). The collected organic layers were dried over sodium sulfate and concentrated. The crude oil was purified by flash chromatography [silica gel, ethyl acetate:hexane (2:8)].

**3.3.1.** (*R*)-2-Acetyl-2-methyl-5-oxo-hexanoic acid ethyl ester 10a. Oil; yield 72%;  $[\alpha]_D = +8.5$  (c = 4.8, CHCl<sub>3</sub>); IR (film, cm<sup>-1</sup>): 1717 1682 1357, 1250; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  1.20 (t, J = 6.0 Hz, 3H), 1.25 (s, 3H), 1.80–2.10 (m, 2H), 2.05 (s, 3H), 2.09 (s, 3H), 2.20–2.45 (m, 2H), 4.1 (q, J = 6.0 Hz, 2H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  207.1 (C), 205.1 (C), 172.4 (C), 61.2 (CH<sub>2</sub>), 58.4 (C), 38.3 (CH<sub>2</sub>), 29.7 (CH<sub>3</sub>), 28.3 (CH<sub>2</sub>), 25.9 (CH<sub>3</sub>), 19.0 (CH<sub>3</sub>), 13.9 (CH<sub>3</sub>). Anal. calcd for C<sub>11</sub>H<sub>18</sub>O<sub>4</sub>: C, 61.66, H 8.47. Found: C, 61.79; H, 8.52%.

**3.3.2.** (*R*)-2-Acetyl-2-ethyl-5-oxo-hexanoic acid ethyl ester 10b. Oil; yield 67%;  $[\alpha]_D = +8.7$  (c = 1.6, CHCl<sub>3</sub>); IR (film, cm<sup>-1</sup>): 2975, 1717 1361, 1235; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  0.7 (t, J = 6.0 Hz, 3H), 1.25 (t, J = 6.0 Hz, 3H), 1.89 (m, 2H), 2.00 (s, 3H), 2.05 (s, 3H), 2.10–2.20 (m, 2H), 2.25–2.40 (m, 2H), 4.2 (q, J = 6.0 Hz, 2H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  209.1 (C), 207.2 (C), 176.0 (C), 60.7 (C), 59.8 (CH<sub>2</sub>), 38.3 (CH<sub>2</sub>), 28.1 (CH<sub>2</sub>), 25.9 (CH<sub>2</sub>), 24.7 (CH<sub>3</sub>), 19.6 (CH<sub>3</sub>), 14.1 (CH<sub>3</sub>), 8.5 (CH<sub>3</sub>). Anal. calcd for C<sub>12</sub>H<sub>20</sub>O<sub>4</sub>: C, 63.14; H, 8.83. Found: C, 63.34; H, 8.72%.

3.3.3. (*S*)-2-Acetyl-2-(3-oxobutyl)-pent-4-enoic acid methyl ester 10c. Oil; yield 73%;  $[\alpha]_D = +18.6$  (c = 9.9, CHCl<sub>3</sub>); IR (film, cm<sup>-1</sup>): 1730, 1714; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  2.05 (s, 3H), 2.07 (s, 3H), 2.05–2.19 (m, 2H), 2.29–2.39 (m, 2H), 2.45–2.65 (m, 2H), 3.65 (s, 3H), 4.90–5.10 (m, 2H), 5.49–5.62 (m, 1H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  207.0 (C), 201.9 (C), 176.8 (C), 131.9 (CH), 119.2 (CH<sub>2</sub>), 62.4 (C), 52.4 (CH<sub>3</sub>), 38.1 (CH<sub>2</sub>), 36.8 (CH<sub>2</sub>), 29.9 (CH<sub>3</sub>), 26.9 (CH<sub>3</sub>), 25.4 (CH<sub>2</sub>). Anal. calcd for C<sub>12</sub>H<sub>18</sub>O<sub>4</sub>: C, 63.70; H, 8.02. Found: C, 63.76; H, 7.79%.

**3.3.4.** (*S*)-2-Acetyl-2-benzyl-5-oxo-hexanoic acid methyl ester 10d. White powder; mp 77–79°C; yield 75%;  $[\alpha]_D = -26.9$  (c = 4.5, CHCl<sub>3</sub>); IR (film, cm<sup>-1</sup>): 1735, 1668; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  2.05 (s, 3H), 2.07 (s, 3H), 2.01–2.10 (m, 2H), 2.15–2.45 (m, 2H), 3.10 (s,

2H), 3.71 (s, 3H), 6.95–7.05 (m, 2H), 7.14–7.20 (m, 3H);  $^{13}$ C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  204.9 (C), 201.3 (C), 171.1 (C), 139.4 (C), 130.7 (2 CH), 128.1 (2 CH), 125.7 (CH), 60.9 (C), 52.3 (CH<sub>3</sub>), 39.7 (CH<sub>2</sub>), 36.1 (CH<sub>2</sub>), 29.4 (CH<sub>2</sub>), 26.4 (CH<sub>3</sub>), 23.1 (CH<sub>3</sub>). Anal. calcd for C<sub>16</sub>H<sub>20</sub>O<sub>4</sub>: C, 69.54; H, 7.30. Found: C, 69.68; H, 7.17%.

# 3.4. General procedure for synthesis of Hagemann's ester cyclisation

To the adduct (0.010 mol) was added piperidine (0.008 mol) and acetic acid (0.0095 mol). The mixture was stirred at 80°C for 1.5 h, dissolved in diethyl ether (20 mL), and the organic phase washed with water (3×15 mL). The aqueous layer was extracted with diethyl ether (2×30 mL). The combined organic extracts were dried over sodium sulfate and concentrated in vacuo and the evaporation residue was purified by flash chromatography [silica gel, ethyl acetate:hexane (2:8)].

- **3.4.1.** (*R*)-1,2-Dimethyl-4-oxo-cyclohex-2-enecarboxylic acid ethyl ester 2a. Oil; yield 78%;  $[\alpha]_D = +108.3$  (c = 4.1, CHCl<sub>3</sub>); IR (film, cm<sup>-1</sup>): 1734, 1678, 1627; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  1.25 (t, J = 7 Hz, 3H), 1.40 (s, 3H), 1.95 (d, J = 1 Hz, 3H), 2.00–2.30 (m, 2H), 2.40–2.60 (m, 2H), 4.20 (q, J = 7 Hz, 2H), 5.90 (q, J = 1 Hz, 1H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  197.8 (C), 173.5 (C), 161.1 (C), 127.7 (CH), 60.9 (CH<sub>2</sub>), 46.9 (C), 33.9 (CH<sub>2</sub>), 33.9 (CH<sub>2</sub>), 21.9 (CH<sub>3</sub>), 19.8 (CH<sub>3</sub>), 13.7 (CH<sub>3</sub>). Anal. calcd for C<sub>11</sub>H<sub>16</sub>O<sub>3</sub>: C, 67.32; H, 8.22. Found: C, 67.20; H, 8.35%.
- 3.4.2. (*R*)-1-Ethyl-2-methyl-4-oxo-cyclohex-2-enecarboxylic acid ethyl ester 2b. Oil; yield 68%;  $[\alpha]_D = +109.7$  (c = 6.2, CHCl<sub>3</sub>); IR (film, cm<sup>-1</sup>): 1714, 1675, 1652;  $^1$ H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  0.86 (t, J = 7 Hz, 3H), 1.26 (t, J = 7 Hz, 3H), 1.65–1.74 (m, 2H) 1.98 (d, J = 1 Hz, 3H), 1.98–2.10 (m, 2H), 2.30–2.60 (m, 2H), 4.18 (q, J = 7 Hz, 2H), 5.95 (q, J = 1 Hz, 1H);  $^{13}$ C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  195.0 (C), 170.5 (C), 162.1 (C), 126.7 (CH), 60.8 (CH<sub>2</sub>), 55.1 (C), 29.5 (CH<sub>2</sub>), 28.0 (CH<sub>2</sub>), 22.9 (CH<sub>2</sub>), 24.8 (CH<sub>3</sub>), 13.9 (CH<sub>3</sub>), 9.8 (CH<sub>3</sub>). Anal. calcd for C<sub>12</sub>H<sub>18</sub>O<sub>3</sub>: C, 68.54; H, 8.63. Found: C, 68.42; H, 8.27%.
- **3.4.3.** (*S*)-1-Allyl-2-methyl-4-oxo-cyclohex-2-enecarboxylic acid methyl ester 2c. Syrup; yield 79%;  $[\alpha]_D = +88.4$  (c = 12.8, CHCl<sub>3</sub>); IR (film, cm<sup>-1</sup>): 1731, 1678; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  1.96 (d, J = 1 Hz, 3H), 2.00–2.07 (m, 1H), 2.25–2.45 (m, 3H), 2.50–2.70 (m, 2H), 3.69 (s, 3H), 5.09–5.15 (m, 2H), 5.60–5.75 (m, 1H), 5.90 (q, J = 1 Hz, 1H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>)  $\delta$  198.1 (C), 173.0 (C), 159.9 (C), 132.8 (CH), 129.4 (CH), 119.4 (CH<sub>2</sub>), 55.7 (C), 50.4 (CH<sub>3</sub>), 39.9 (CH<sub>2</sub>), 34.1 (CH<sub>2</sub>), 30.4 (CH<sub>2</sub>), 21.3 (CH<sub>3</sub>). Anal. calcd for C<sub>12</sub>H<sub>16</sub>O<sub>3</sub>: C, 69.21; H, 7.74. Found: C, 69.18; H, 7.77%.
- **3.4.4.** (*S*)-1-Benzyl-2-methyl-4-oxo-cyclohex-2-enecarboxylic acid methyl ester 2d. White powder; mp 40–43°C; yield 65%; [ $\alpha$ ]<sub>D</sub>=-62.5 (c=3.2, CCl<sub>4</sub>); IR (film, cm<sup>-1</sup>): 3100, 1740, 1680; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$

1.78–1.95 (m, 1H), 2.0 (d, J=1 Hz, 3H), 2.10–2.50 (m, 3H), 2.80 (d, J=14 Hz, 1H), 3.45 (d, J=14 Hz, 1H), 3.70 (s, 3H), 5.9 (d, J=1 Hz, 1H), 7.0–7.1 (m, 2H), 7.15–7.25 (m, 3H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>) δ 194.6 (C), 171.4 (C), 162.2 (C), 136.5 (C), 130.7 (2 CH), 128.1 (2 CH), 126.7 (CH), 126.0 (CH), 56.9 (C), 52.4 (CH<sub>3</sub>), 39.8 (CH<sub>2</sub>), 29.5 (CH<sub>2</sub>), 28.8 (CH<sub>2</sub>), 24.1 (CH<sub>3</sub>). Anal. calcd for C<sub>16</sub>H<sub>18</sub>O<sub>3</sub>: C, 74.39; H, 7.02. Found: C, 74.52; H, 7.29%.

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